

METABOLISM OF CEPHALEXIN-¹⁴C* IN MICE AND IN RATS

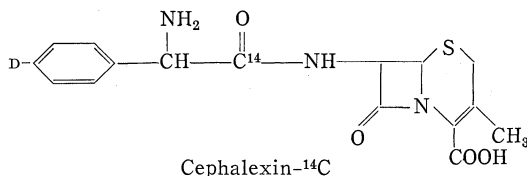
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The metabolic fate of the orally effective antibiotic, cephalixin, has been studied in the rat and the mouse. Cephalixin is efficiently absorbed from the gastrointestinal tract as the intact antibiotic. Cephalixin is not metabolized in the body and is eliminated as unaltered antibiotic primarily via the urine. Cephalixin is more rapidly absorbed and eliminated by the mouse than by the rat.

Cephalixin is a broad spectrum antibiotic, active against gram positive cocci, including penicillin-resistant strains, as well as against many gram-negative organisms¹⁾. Cephalixin, a cephalosporin antibiotic, structurally related to D-cephaloglycin^{2)*}, appears to be efficiently absorbed from the gastrointestinal tract in both laboratory animals^{3,3)} and in humans⁴⁾. This antibiotic also appears to be unusually resistant to metabolic alteration^{3,4)}. As in earlier studies on the fate of cephalosporin antibiotics *in vivo* from these laboratories^{5,6,7)}, radiocarbon labeling was used to facilitate the investigation.



Materials and Methods

D-(-)-2-Phenylglycine-¹⁴C. The procedure used for the preparation of racemic 2-phenylglycine-¹⁴C and the enzymatic resolution into the enantiomeric forms has been described by BILLINGS and SULLIVAN⁸⁾.

Cephalixin-¹⁴C. D-Phenylglycine-¹⁴C (3.05 m-moles, 11.56 μ c/mg) was allowed to react with 3.1 m-moles of methyl acetoacetate in a solution consisting of 3.1 m-moles of sodium hydroxide, 0.15 ml water and 1.2 ml of methanol. After heating for 30 minutes at 60°C, the clear reaction solution was evaporated to dryness *in vacuo*. The residual oil was dried for 24 hours *in vacuo* over concentrated sulfuric acid to yield semi-solid methyl 3-(D- β -carboxybenzylamino)crotonate-¹⁴C sodium salt.

Ethyl chloroformate (3.7 m-moles) was allowed to react with a solution consisting of 3.1 m-moles of methyl 3-(D- α -carboxybenzylamino)crotonate-¹⁴C sodium salt in 9.2 ml of

* Cephalixin is the generic name given to 7-(D-2-amino-2-phenylacetamido)-3-methyl-3-cephem-4-carboxylic acid. D-Cephaloglycin is the generic name given to 7-(D-2-amino-2-phenylacetamido)-cephalosporanic acid. D- α -Aminophenyl[1-¹⁴C]acetic acid will be designated as D-2-phenylglycine-¹⁴C.

acetonitrile containing one drop of N,N-dimethylbenzylamine at -10°C . After strring the solution for 15 minutes at this temperature, a solution of 670 mg (3.0 m-moles) of 7-amino-desacetoxycephalosporanic acid (7-ADCA)⁹⁾ in 8 ml of acetonitrile-water (1 : 1 v/v) containing 3.0 m-moles of triethylamine was added rapidly with stirring. After 30 minutes 1.0 g of sodium chloride was added to the reaction mixture and stirring was continued for 15 minutes at 0°C . The organic phase was separated and to it was added a solution of 0.35 ml of 100 % formic acid and 0.11 ml of water. The resulting solution was stirred at 0°C for 1 hour to facilitate crystallization of cephalixin- ^{14}C . The product was collected by filtration, washed with 5.0 ml of acetonitrile and dried *in vacuo*. The yield was 165 mg (16 %) of pure cephalixin- ^{14}C with a specific radioactivity of $4.90 \mu\text{c}/\text{mg}$. Paper chromatography followed by biological and radiological assay showed the product to be pure.

Chromatographic Methods. A combination of paper and thin-layer chromatography was employed for the detection of radioactive metabolites. Paper chromatography was carried out on Whatman #1 paper and thin-layer chromatography (TLC) was carried out on Eastman Cellulose Chromagram sheets. Areas of biological activity were located by the bioautographic method described by MILLER¹⁰⁾ using *Sarcina lutea*. The radioactive zones were located by means of a scanner (Vanguard 880 Automatic Chromatogram Scanner) and by radioautography using Kodak Medical X-Ray film.

Paper chromatograms, developed with butan-1-ol - acetic acid - water (3 : 1 : 1 by volume) were used for the identification and quantitation of radioactive metabolites. Radiocarbon quantitation was accomplished by the uniform sectioning of the paper chromatogram and subsequent liquid scintillation counting. Results obtained with the above system were confirmed by the use of one or more of the following systems :

1. TLC employing a butan-1-ol - acetic acid - water (3 : 1 : 1) solvent system.
2. Paper chromatography employing an ethyl acetate - acetic acid - water (3 : 1 : 1) solvent system.
3. TLC employing an ethyl acetate - acetic acid - water (3 : 1 : 1) solvent system.
4. TLC employing an acetonitrile - ethyl acetate - water (3 : 1 : 1) solvent system.

Urinary, Fecal and Biliary Excretion Studies. An aqueous solution of cephalixin- ^{14}C , 1.6 mg/ml, was administered by the oral and intraperitoneal routes ($46 \mu\text{moles}/\text{kg}$) to groups of 200 g male Purdue-Wistar rats and 30 g I. C. R. (Cox) mice. The test animals were kept in stainless steel metabolism cages. Urine samples were collected at regular intervals (2 hours) and feces samples were collected after 24 hours.

The radioactivity contained in each urine sample was determined by liquid scintillation counting. The nature and quantitation of the radioactive metabolites in urine was determined by paper and thin-layer chromatographic analysis as described above. The biologically active metabolite was identified by comparison chromatography using an authentic sample of cephalixin- ^{14}C .

Feces samples were dried, ground and analyzed for radiocarbon content by the wet tissue digestion method described by MAHIN and LOFBERG¹¹⁾.

For the bile study, 200 g male rats were anesthetized with ether and a cannula placed into the common bile duct. The cephalixin- ^{14}C was administered orally and the bile fluid collected for 24 hours. Radiocarbon content was determined by liquid scintillation counting of an aliquot of the bile fluid.

Blood and Tissue Level Studies. An aqueous solution of cephalixin- ^{14}C ($46 \mu\text{moles}/\text{kg}$) was administered orally and intraperitoneally to separate groups of three 200 g Purdue-Wistar rats. At given time intervals blood samples were removed from the tail vein. The samples, averaging 200 mg in weight, were processed for liquid scintillation counting using the wet tissue digestion method of MAHIN and LOFBERG¹¹⁾.

Eighteen, 30 g male I. C. R. (Cox) mice were administered orally an aqueous solution of cephalixin- ^{14}C ($46 \mu\text{moles}/\text{kg}$). At six time intervals, 3 mice were sacrificed by decapitation and the blood was collected. The blood of each mouse was sampled and was

processed for liquid scintillation counting by the wet tissue digestion method.

For the tissue level determinations, cephalixin- ^{14}C (46 $\mu\text{moles/kg}$) was administered orally to 6 male Harlan rats and 6 I. C. R. (Cox) mice. One and four hours after administration, a group of three animals was sacrificed by decapitation. The appropriate tissues and organs were removed and their wet weights recorded. A weighed sample of each tissue and organ was removed and was prepared for liquid scintillation counting employing the wet tissue digestion method.

Results and Discussion

Excretion Studies. Rat

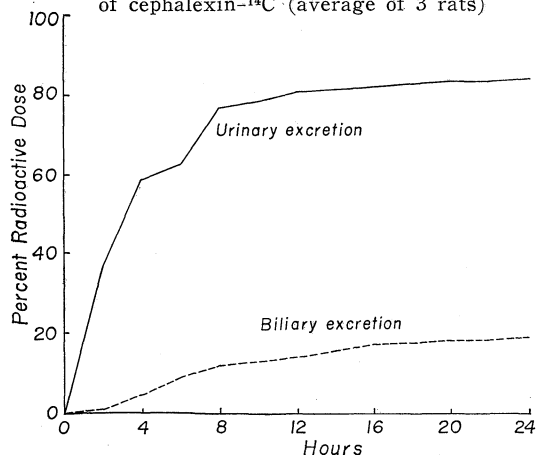
Fig. 1 shows the recovery of total urinary radioactivity as well as the distribution of radioactive metabolites in the urine of rats following the oral administration of a single dose of cephalixin- ^{14}C . Sixty percent of the administered radiocarbon was recovered in the urine after 4 hours, 77% after 8 hours and 84% after 24 hours. The feces, collected over this period of time, were found to contain 15% of the administered radiocarbon. Cannulation of the common bile duct of a rat immediately prior to the oral administration of a single dose of cephalixin- ^{14}C and subsequent collection of a 24-hour sample of the bile fluid, revealed that 18.2% of the administered radiocarbon was eliminated via biliary excretion (Fig. 1). This result suggests that the greater part of the radioactivity found in the feces of the normal rat was due to biliary excretion and not to the incomplete adsorption of the cephalixin.

The individual urine and bile samples were examined for the presence of biologically active and radioactive metabolites using paper and thin-layer chromatographic techniques. The results of these examinations revealed the presence of only one metabolite. It was both biologically active and radioactive. This biologically active ^{14}C -labeled metabolite was identified as unchanged cephalixin- ^{14}C by chromatographic comparison with authentic cephalixin- ^{14}C . The presence of any possible metabolite arising from the hydrolytic cleavage of the side chain amide bond⁷⁾, *i. e.* 2-phenylglycine- ^{14}C , benzoylformic acid- ^{14}C , could not be demonstrated.

Excretion Studies. Mice

The oral administration of cephalixin- ^{14}C to a group of I. C. R. (Cox) mice resulted in the rapid elimination of a large percentage of the administered radiocarbon via the kidney. Sixty-two percent of the administered radioactivity appeared in the urine in 3 hours, 80% in 7 hours and 89% in 24 hours. Chromatographic examination of

Fig. 1. Total urinary and biliary radioactivity recoveries following oral administration of cephalixin- ^{14}C (average of 3 rats)



these urine fractions again showed that the only radioactive entity present was unchanged cephalixin.

These results, obtained from excretion studies in rats and mice, show cephalixin to be completely absorbed from the gastrointestinal tract following oral administration. The recovery of unaltered ^{14}C -labeled antibiotic as the single radioactive component present in the urine and the bile fluid after oral administration demonstrates the *in vivo* stability of cephalixin. Although cephalixin is structurally related to cephaloglycin, it is more efficiently absorbed in the gastrointestinal tract than cephaloglycin. Cephalixin is resistant to enzymatic hydrolysis of the side-chain amide moiety, an important metabolic pathway in the biodegradation of *d*-cephaloglycin.

Whole Blood Radiocarbon Levels

The radiocarbon blood level curves obtained after intraperitoneal and oral administration of 46 $\mu\text{moles/kg}$ of cephalixin- ^{14}C to rats are shown in Fig. 2. A maximum radiocarbon level equivalent to 9.0 μg of cephalixin- ^{14}C per ml of blood was obtained 30 minutes after intraperitoneal administration. Radiocarbon levels declined with the half-life of about 1.5 hours. Because of *in vivo* stability of cephalixin it is reasonable to assume that this figure represents the half-life of cephalixin in rats. The peak blood level of cephalixin- ^{14}C following oral administration was 3.8 $\mu\text{g/ml}$ of blood. This was attained 1 hour after administration. The half-life of cephalixin after oral administration calculated from these results was also near 1.5 hours. The long term blood level (24 hours) curves in each instance did not appear to be bimodal in character.

The radiocarbon blood level curve obtained after oral administration of 46 $\mu\text{moles/kg}$ of cephalixin- ^{14}C in mice is shown in Fig. 3. A peak level of 6 $\mu\text{g/ml}$ of blood

Fig. 2. Whole blood radiocarbon levels in rats after oral and intraperitoneal administration of cephalixin- ^{14}C (average of 3 rats)

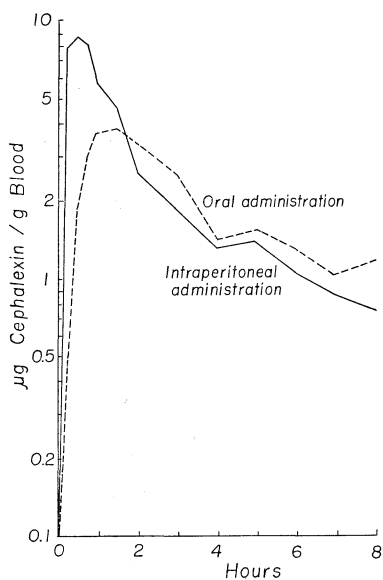
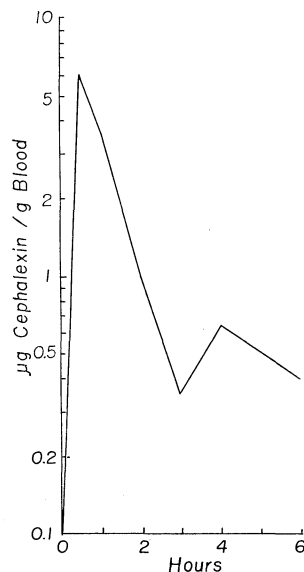


Fig. 3. Whole blood radiocarbon levels in mice following intraperitoneal administration of cephalixin- ^{14}C (average of 3 mice)



was attained 30 minutes after administration. The half-life of cephalixin-¹⁴C in mice estimated from this data is 45 minutes.

Tissue Level Studies

Cephalixin-¹⁴C was administered orally to rats and mice. The radiocarbon levels in tissue and organs were determined 1 and 4 hours after administration. Since the urinary and biliary excretion studies showed that all of the administered cephalixin-¹⁴C was eliminated unchanged via the bile duct and the kidney, the level of radiocarbon in the tissues can be expressed as μg of cephalixin per g of body tissue. Table 1 shows the various tissue levels of cephalixin-¹⁴C at 1 and 4 hours after oral administration of 46 $\mu\text{moles/kg}$ of the labeled antibiotic.

The results of this study show that while all tissues examined contained

some cephalixin-¹⁴C, the only tissues possessing higher levels than that of the blood were the kidney and the liver, organs responsible for its elimination. It is of interest to note that the mouse eliminates the antibiotic at a rate faster than that of the rat.

Table 1. Cephalixin-¹⁴C tissue levels in rats and in mice after a single oral dose of cephalixin-¹⁴C (46 $\mu\text{moles/kg}$)

Tissue	μg Cephalixin/g tissue			
	Rat		Mouse	
	1 hr.	4 hrs.	1 hr.	4 hrs.
Blood	3.71	2.09	3.59	0.53
Liver	17.11	7.25	12.96	1.93
Spleen	2.21	1.45	1.45	0.40
Kidney	39.93	23.69	27.23	3.53
Lung	3.38	2.58	1.63	0.30
Heart	1.52	1.09	3.31	1.07
Fat	1.54	0.80	1.41	0.34
Muscle	1.16	0.76	1.11	0.32
Brain	0.53	0.24	0.30	0.11

Summary

The study of the metabolic fate of cephalixin-¹⁴C in the rat and the mouse has shown this antibiotic to be quantitatively absorbed in the gastrointestinal tract as the intact antibiotic. Cephalixin is not metabolized in the body and is eliminated as the unchanged antibiotic via the kidney. Cephalixin is more rapidly absorbed and eliminated by the mouse than by the rat.

These results are of particular interest when compared to those obtained from the study of the metabolism of *d*-cephaloglycin⁷⁾. Cephaloglycin is metabolized in the rat by two pathways: hydrolysis of the amide linkage to form *D*-2-phenylglycine and by deacetylation to form desacetylcephaloglycin. Thus the replacement of the acetoxymethyl moiety of cephaloglycin by a methyl group produces an antibiotic, cephalixin, which is remarkably more resistant to metabolism and which is more readily absorbed from the gut of the rat and the mouse.

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